



# ESCAPING SPIN WIZARDRY IN PAIN MEDICINE

LEARNING CLASSIFICATION, QUANTIFICATION, AND SPECIFICATION (2025)



**Presenter:** Guilherme (Gui) Dos Santos, MD | **Department of Anesthesiology, Reanimation, and Pain Medicine**

Hospital Clínic de Barcelona, University of Barcelona

# CONFLICT OF INTEREST DISCLOSURE

No actual or potential financial conflicts of interest associated with the material presented in this lecture.

## SUMMARY

- A) Learning Points for the Session.
- B) Types of PRP Concentrates and the Importance of Leukocytes.
- C) Influence of Outside Elements in the Preparation.
- D) The Importance of Interpersonal Variations in Whole Blood Samples.

## SUMMARY

- E) Biochemical Effects of PRP Concentrates in Different Target Tissues (Joint and Tendon).
- F) The Importance of Classification, Quantification, and Specification.
- G) Controversies and Future Directions of Research.

## LEARNING POINTS BY THE END OF THE SESSION:

- A) **DISTINGUISH** between different types of PRP Concentrates.
- B) **UNDERSTAND** the potential influence of external elements in the PRP concentrate.
- C) **ENUMERATE** the main biological effects of PRP in different target tissues (joint and tendon).
- D) **RECOGNIZE** the clinical importance of classification, quantification, and specification.

## Daring discourse

# From spin wizardry to regenerative alchemy: a philosophical inquiry into healing and standardization

Guilherme Ferreira-Dos-Santos  1,2

product, may only observe modest improvement. These concrete examples of variability serve as poignant reminders that our current nomenclature often masks profound methodological differences.

The ancient parable of the blind men and the elephant offers a striking metaphor here. Each blind man, touching only a part of the elephant, describes it in entirely different terms—a snake, a

Pain Physician 2022; 25:15-27 • ISSN 1533-3159

**Narrative Review**

# **Autologous Platelet-Rich Plasma Applications in Chronic Pain Medicine: Establishing a Framework for Future Research - A Narrative Review**

Guilherme Ferreira-Dos-Santos, MD<sup>1</sup>, Mark Friedrich B. Hurdle, MD<sup>2</sup>,  
Steven R. Clendenen, MD<sup>3</sup>, Jason S. Eldrige, MD<sup>2</sup>, and Wenchun Qu, MD, PhD<sup>2</sup>

Table 1. *The most important α-granule growth factors and their contributions to the process of tissue healing and repair.*

| Growth Factor | Role in Tissue Healing and Repair                                                                                                                                                                                                                                                                                                                                                                                                 |
|---------------|-----------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|
| bFGF-1        | <ol style="list-style-type: none"> <li>1. May contribute to stimulate angiogenesis;</li> <li>2. Mediates cell migration;</li> <li>3. Stimulates proliferation of capillary endothelial cells;</li> <li>4. Influences fibroblasts to produce collagenase;</li> <li>5. Contributes to the production of granulation tissue.</li> </ol>                                                                                              |
| EGF           | <ol style="list-style-type: none"> <li>1. Mitogenic factor;</li> <li>2. Stimulates the proliferation and differentiation of fibroblasts, epidermal and epithelial cells;</li> <li>3. May play an enhancer role on osteogenic differentiation of mesenchymal stem cells (by increasing ECM mineralization).</li> </ol>                                                                                                             |
| HGF           | <ol style="list-style-type: none"> <li>1. Mitogenic factor;</li> <li>2. Promotes angiogenesis (particularly after ischemia);</li> <li>3. Accelerates healing by promoting the dedifferentiation of epidermal cells;</li> <li>4. Regulates cell growth, cell motility, and morphogenesis.</li> </ol>                                                                                                                               |
| IGF-1         | <ol style="list-style-type: none"> <li>1. Expressed mainly in the early inflammatory phase;</li> <li>2. Anabolic effects;</li> <li>3. Stimulates protein synthesis, the proliferation of myoblasts and fibroblasts;</li> <li>4. Enhances collagen and ECM matrix;</li> <li>5. May contribute to early modulation of edema.</li> </ol>                                                                                             |
| PDGF-AB       | <ol style="list-style-type: none"> <li>1. Expressed in the early stages of tendon repair;</li> <li>2. Facilitates the proliferation of other growth factors;</li> <li>3. Attracts mesenchymal stem cells and leukocytes;</li> <li>4. Stimulates angiogenesis;</li> <li>5. Contributes to tissue remodeling.</li> </ol>                                                                                                            |
| TGF-β1        | <ol style="list-style-type: none"> <li>1. Proinflammatory type 2 cytokine (antibody response enhancer);</li> <li>2. Immunosuppressant during the early inflammatory phase;</li> <li>3. Aids in cell migration and fibronectin binding;</li> <li>4. Augments production of tendon sheath fibroblasts;</li> <li>5. Improves tendon mechanics during the healing process;</li> <li>6. Controls angiogenesis and fibrosis.</li> </ol> |
| VEGF          | <ol style="list-style-type: none"> <li>1. Expression peaks in the late inflammatory and proliferative phases;</li> <li>2. Promotes angiogenesis (neovascularization).</li> </ol>                                                                                                                                                                                                                                                  |

Legend: bFGF-1, Basic fibroblast growth factor one; ECM, Extracellular matrix; EGF, Epidermal growth factor; HGF, Hepatocyte growth factor; IGF-1, Insulin-like growth factor 1; PDGF-Aβ, Platelet-derived growth factor AB; TGF-β1, Transforming growth factor β1; VEGF, Vascular endothelial growth factor.

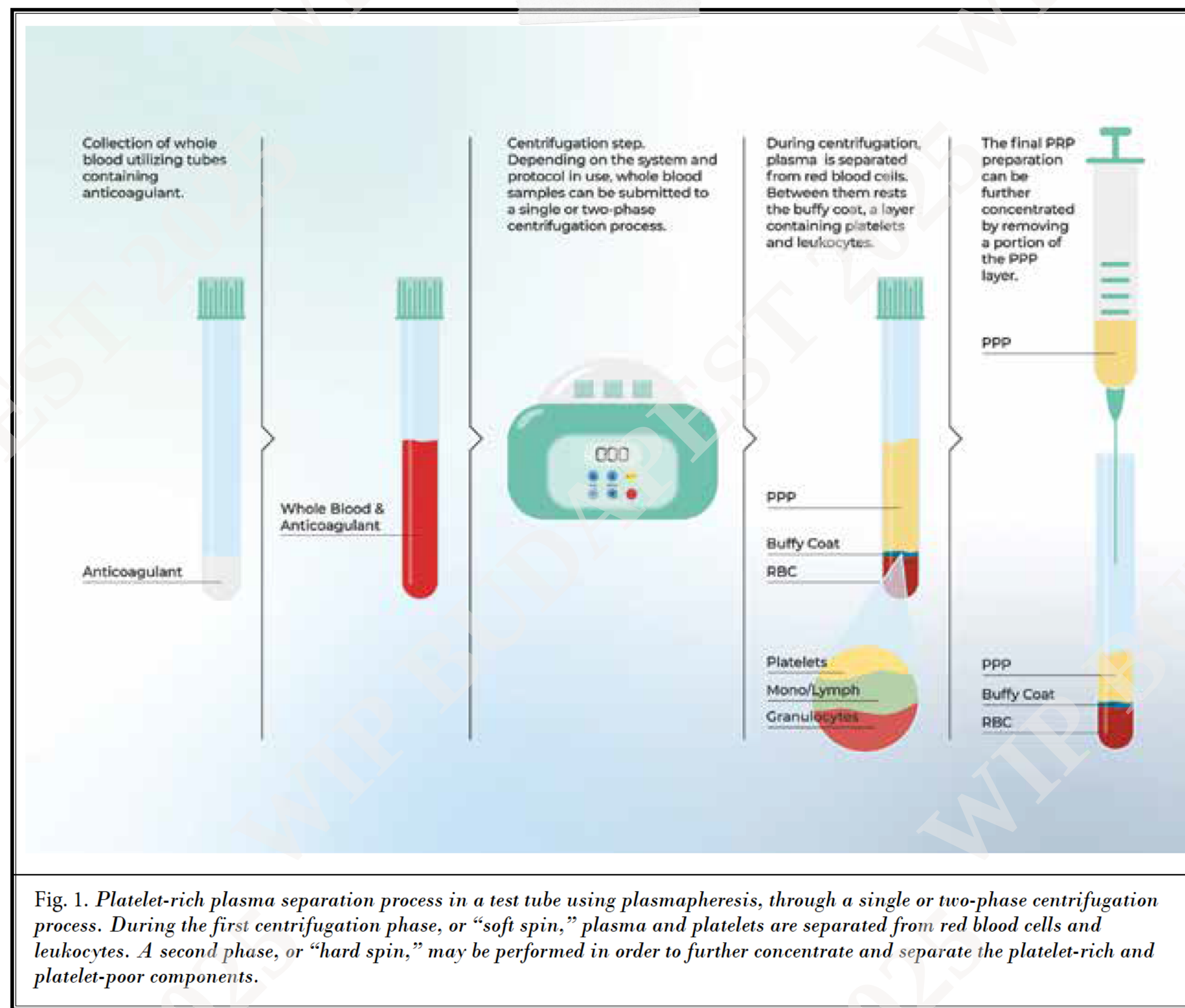
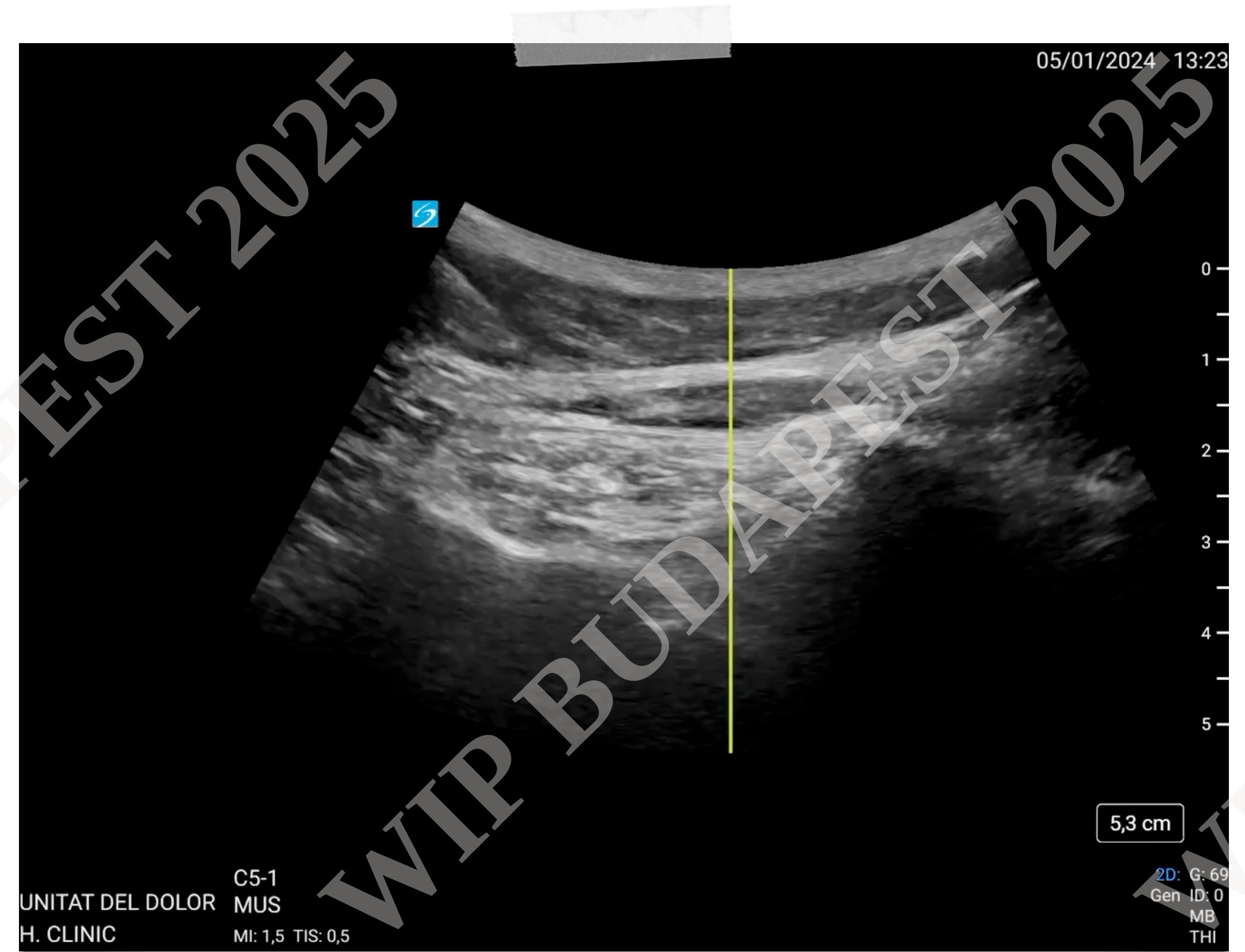


Fig. 1. Platelet-rich plasma separation process in a test tube using plasmapheresis, through a single or two-phase centrifugation process. During the first centrifugation phase, or “soft spin,” plasma and platelets are separated from red blood cells and leukocytes. A second phase, or “hard spin,” may be performed in order to further concentrate and separate the platelet-rich and platelet-poor components.



# TYPES OF PRP CONCENTRATES AND THE IMPORTANCE OF LEUKOCYTES

Because the inclusion of leukocytes in the final preparation may affect the potency and efficacy of the final product, the more generic term “PRP” does not distinguish between different products with different specificities.

Wasterlain AS, Braun HJ, Dragoo JL. **Contents and Formulation of Platelet-rich Plasma.** *Operative Tech Orthop.* 2012;22:33-42.

Kobayashi Y, Saita Y, Nishio H, *et al.* **Leukocyte Concentration and Composition in Platelet-rich Plasma (PRP) Influences the Growth Factor and Protease Concentrations.** *J Orthop Sci.*

2016;21:683-689.

# TYPES OF PRP CONCENTRATES AND THE IMPORTANCE OF LEUKOCYTES

**Leukocyte-rich PRP (LR-PRP)** - PRP Preparation with leukocyte count **ABOVE** the patient's baseline. Associated with a more intense **PRO-INFLAMMATORY** local response.

**Leukocyte-poor PRP (LP-PRP)** - PRP preparation with leukocyte count **BELOW** the patient's baseline. Mainly associated with an **ANTI-INFLAMMATORY** local response.

Wasterlain AS, Braun HJ, Dragoo JL. **Contents and Formulation of Platelet-rich Plasma**. *Operative Tech Orthop*. 2012;22:33-42.

Kobayashi Y, Saita Y, Nishio H, *et al*. **Leukocyte Concentration and Composition in Platelet-rich Plasma (PRP) Influences the Growth Factor and Protease Concentrations**. *J Orthop Sci*.

2016;21:683-689.

# TYPES OF PRP CONCENTRATES AND THE IMPORTANCE OF LEUKOCYTES

**Leukocyte-rich PRP (LR-PRP)** - Induces predominantly **CATABOLIC** and **INFLAMMATORY** changes, such as increased MMP 1, MMP-13, IL-1 $\beta$ , IL-6, and TNF- $\alpha$ .

**Leukocyte-poor PRP (LP-PRP)** - Induces predominantly **ANABOLIC** effects such as increased gene expression of anabolic genes, alpha-smooth muscle actin, and collagen types I and III.

# INFLUENCE OF OUTSIDE ELEMENTS IN THE PREPARATION

**The initial whole blood sample is most often collected in the presence of an anticoagulant, which binds calcium and prevents the initiation of the clotting cascade through inhibition of the conversion of prothrombin to thrombin.**

## INFLUENCE OF OUTSIDE ELEMENTS IN THE PREPARATION

Table 2. Anticoagulants utilized for platelet-rich plasma concentrate preparation and their effects on platelet count and recovery, morphology, platelet morphology,  $\alpha$ -granule growth factor release, and bone marrow-derived mesenchymal stromal cell gene expression.

| Anticoagulant  | Platelet Count / Recovery Rate       | Platelet Morphology                                 | Growth Factor Release                                               | BM-MSK Gene Expression           |
|----------------|--------------------------------------|-----------------------------------------------------|---------------------------------------------------------------------|----------------------------------|
| ACD-A          | Lowest average count / 45% of total  | No influence on MPV                                 | No significant differences in TGF $\beta$ -1 and VEGF concentration | 30% different from control group |
| EDTA           | Highest average count / 76% of total | MPV increase, likely indicating platelet activation |                                                                     | 47% different from control group |
| Sodium Citrate | Medium average count / 81% of total  | No influence on MPV                                 |                                                                     | 25% different from control group |

Legend: ACD-A, Anticoagulant citrate dextrose solution A; BM-MSK, Bone marrow-derived mesenchymal stromal cells; EDTA, ethylenediamine-tetraacetic acid; MPV, Mean platelet volume; TGF- $\beta$ 1, Transforming growth factor  $\beta$ 1; VEGF, Vascular endothelial growth factor.

# INFLUENCE OF OUTSIDE ELEMENTS IN THE PREPARATION

Some studies have also reported the utilization of PRP concentrates prepared in the absence of an anticoagulant. **In such circumstances, the time required between whole blood collection and injection must be substantially shortened.**

# INFLUENCE OF OUTSIDE ELEMENTS IN THE PREPARATION

**PRP separation protocols also vary in platelet activation method. ACTIVATION** refers to 2 key processes: The **DEGRANULATION OF PLATELETS** to release growth factors from their  $\alpha$ -granules, and the **CLEAVAGE OF FIBRINOGEN** to initiate matrix formation.

# INFLUENCE OF OUTSIDE ELEMENTS IN THE PREPARATION

**Several of the commercial kits available on the market already include an activation step before PRP administration**, commonly by adding thrombin and/or calcium chloride. However, external activation may not be necessary, as PRP activates when in contact with fibrillar collagen.

Wasterlain AS, Braun HJ, Dragoo JL. **Contents and Formulation of Platelet-rich Plasma**. *Operative Tech Orthop*. 2012;22:33-42.

Kobayashi Y, Saita Y, Nishio H, *et al*. **Leukocyte Concentration and Composition in Platelet-rich Plasma (PRP) Influences the Growth Factor and Protease Concentrations**. *J Orthop Sci*.

2016;21:683-689.

## INFLUENCE OF OUTSIDE ELEMENTS IN THE PREPARATION

Table 3. *Head-to-head comparison of the protocols for platelet-rich plasma separation from 3 different commercial systems.*

| Commercial Systems                                | Whole Blood Volume (mL) | Anticoagulant Utilized | Centrifuge Force (g) / Centrifuge Time | Final Volume of PRP (mL) |
|---------------------------------------------------|-------------------------|------------------------|----------------------------------------|--------------------------|
| Cascade (MTF Sport Medicine, Edison, New Jersey)  | 18                      | Sodium citrate, 2 mL   | 1100 / 6 min                           | 7.5                      |
| GPS III (Biomet Orthopedics Inc, Warsaw, Indiana) | 55                      | ACD-A, 5 mL            | 1100 / 15 min                          | 6.0                      |
| Magellan (Arteriocyte Inc, Cleveland, Ohio)       | 26                      | ACD-A, 5 mL            | 1200 / 17 min                          | 6.0                      |

Legend: ACD-A, Anticoagulant citrate dextrose solution A; PRP, Platelet-rich plasma.

## INFLUENCE OF OUTSIDE ELEMENTS IN THE PREPARATION

Table 4. *Comparison of mean platelet concentration and platelet capture efficiency, red blood cell, white blood cell, and fibrinogen concentrations between three different platelet-rich plasma separation systems.*

| Commercial Systems                                | Platelet Concentration (x103/ $\mu$ L) | Platelet Capture Efficiency Rate (%) | Red Blood Cells (x103/ $\mu$ L) | White Blood Cells (x103/ $\mu$ L) | Fibrinogen (mg/dL) |
|---------------------------------------------------|----------------------------------------|--------------------------------------|---------------------------------|-----------------------------------|--------------------|
| Cascade (MTF Sport Medicine, Edison, New Jersey)  | 443.8 $\pm$ 24.7                       | 67.6 $\pm$ 4.1                       | 0.1 $\pm$ 0.1                   | 1.1 $\pm$ 0.2                     | 283.8 $\pm$ 34.2   |
| GPS III (Biomet Orthopedics Inc, Warsaw, Indiana) | 566.2 $\pm$ 292.6                      | 22.6 $\pm$ 11.8                      | 1.5 $\pm$ 1.7                   | 34.4 $\pm$ 13.6                   | 286.0 $\pm$ 42.7   |
| Magellan (Arteriocyte Inc, Cleveland, Ohio)       | 780.2 $\pm$ 246.5                      | 65.5 $\pm$ 19.6                      | 0.5 $\pm$ 0.3                   | 11.0 $\pm$ 8.2                    | 277.4 $\pm$ 30.5   |

Adapted from: Castillo TN, Pouliot MA, Kim HJ, Dragoo JL. **Comparison of Growth Factor and Platelet Concentration from Commercial Platelet-rich Plasma Separation Systems.** *Am J Sports Med.* 2011;39(2):266-271.

# INFLUENCE OF OUTSIDE ELEMENTS IN THE PREPARATION

Table 5. Comparison of mean growth factor concentrations between 3 different platelet-rich plasma separation systems.

| Commercial Systems                                | PDGF- $\alpha\beta$<br>(ng/mL) | PDGF- $\beta\beta$<br>(ng/mL) | TGF- $\beta$ 1<br>(ng/mL) | VEGF<br>(ng/mL) |
|---------------------------------------------------|--------------------------------|-------------------------------|---------------------------|-----------------|
| Cascade (MTF Sport Medicine, Edison, New Jersey)  | 9.7 $\pm$ 3.6                  | 14.8 $\pm$ 2.5                | 0.1 $\pm$ 0.08            | 0.3 $\pm$ 0.3   |
| GPS III (Biomet Orthopedics Inc, Warsaw, Indiana) | 18.7 $\pm$ 12.8                | 23.1 $\pm$ 10.1               | 0.1 $\pm$ 0.08            | 2.4 $\pm$ 1.1   |
| Magellan (Arteriocyte Inc, Cleveland, Ohio)       | 34.4 $\pm$ 10.7                | 33.0 $\pm$ 8.2                | 0.2 $\pm$ 0.1             | 1.2 $\pm$ 0.8   |

Legend: PDGF- $\alpha\beta$ , Platelet-derived growth factor  $\alpha\beta$ ; PDGF- $\beta\beta$ , Platelet-derived growth factor  $\beta\beta$ ; TGF- $\beta$ 1, Transforming growth factor  $\beta$ 1; VEGF, Vascular endothelial growth factor.

# INTERPERSONAL VARIATIONS IN WHOLE BLOOD SAMPLES

The normal range of platelets in the whole blood of a healthy individual is 150,000 to 350,000 platelets/ $\mu\text{L}$ . **Past consensus agreed on a working definition of platelet concentrate as a preparation with a MINIMUM of  $1.0 \times 10^6$  platelets/ $\mu\text{L}$  ( $1.0 \times 10^9$  platelets/mL).**

Marx RE. **Platelet-rich Plasma (PRP): What is PRP and what is not PRP?**. *Implants Dental*. 2001;10:225-228.

Weibrich G, Hansen T, Kleis W, Buch R, Hitzler WE. **Effect of Platelet Concentration in Platelet-rich Plasma on Peri-implant Bone Regeneration**. *Bone*. 2004;34:665-671.

# INTERPERSONAL VARIATIONS IN WHOLE BLOOD SAMPLES

In other words, for a preparation to be classified as PRP, the respective concentration of platelets should be **INCREASED BY 3 TO 5 FOLD** over the normal baseline.

Marx RE. **Platelet-rich Plasma (PRP): What is PRP and what is not PRP?**. *Implants Dental*. 2001;10:225-228.

Weibrich G, Hansen T, Kleis W, Buch R, Hitzler WE. **Effect of Platelet Concentration in Platelet-rich Plasma on Peri-implant Bone Regeneration**. *Bone*. 2004;34:665-671.

# INTERPERSONAL VARIATIONS IN WHOLE BLOOD SAMPLES

**Variations in platelet concentrations among individuals, in addition to the daily variation in platelet parameters observed within individuals, can further affect the consistency, efficacy, and clinical outcomes of the final product.**

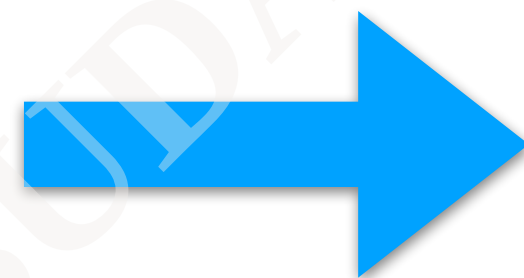
# BIOCHEMICAL EFFECTS OF PRP CONCENTRATES IN DIFFERENT TISSUES

When cultured with tendon stem cells (TSC) isolated from the healthy patellar tendons of rabbits, both **LR-PRP** and **LP-PRP** induced similar TSC differentiation into active tenocytes.

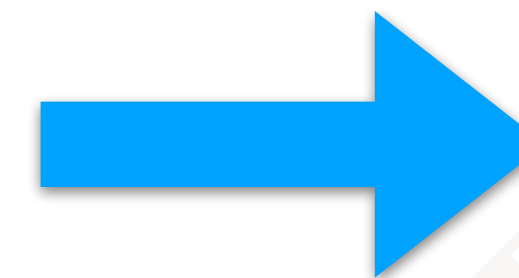
However, **LR-PRP** induced mainly **CATABOLIC** and **INFLAMMATORY** changes in differentiated tenocytes, while **LP-PRP** induced predominantly **ANABOLIC** effects.

# BIOCHEMICAL EFFECTS OF PRP CONCENTRATES IN DIFFERENT TISSUES

**LR-PRP**



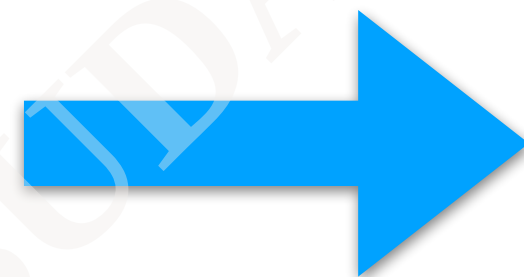
**CATABOLIC and PRO-INFLAMMATORY**



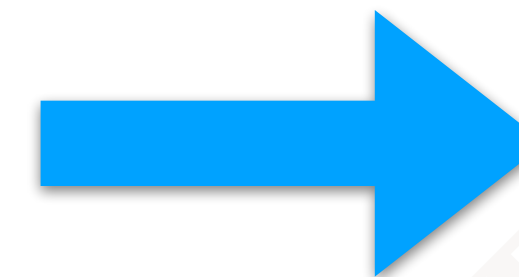
**CHRONIC TENDINOPATHY**

# BIOCHEMICAL EFFECTS OF PRP CONCENTRATES IN DIFFERENT TISSUES

**LP-PRP**



**ANABOLIC and ANTI-INFLAMMATORY**



**CHRONIC TENDINOPATHY**



# BIOCHEMICAL EFFECTS OF PRP CONCENTRATES IN DIFFERENT TISSUES

Although in the acute phase, **LR-PRP** may exert detrimental effects due to its **CATABOLIC** activity, the same **CATABOLIC** activity coupled with the **PRO-INFLAMMATORY** changes appears vital in its role in the treatment of chronic tendinopathy.

# BIOCHEMICAL EFFECTS OF PRP CONCENTRATES IN DIFFERENT TISSUES

Conversely, the use of **LP-PRP** in soft tissue injury may result in **EXCESSIVE SCAR FORMATION** due to the **strong potential** of inducing **INORDINATE ANABOLIC EFFECTS**, especially when **administered in the acute phase**.

# BIOCHEMICAL EFFECTS OF PRP CONCENTRATES IN DIFFERENT TISSUES

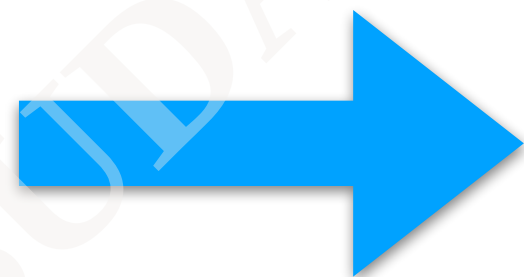
When cultured with synovial fibroblasts isolated from patients with osteoarthritis, **LR-PRP** induced a greater **INCREASE** in **PRO-INFLAMMATORY** IL-1 $\beta$ , IL-8, and bFGF-2, while **DECREASING ANTI-CATABOLIC** factors in chondrocytes, such as HGF and tissue inhibitor of MMP-4.

# BIOCHEMICAL EFFECTS OF PRP CONCENTRATES IN DIFFERENT TISSUES

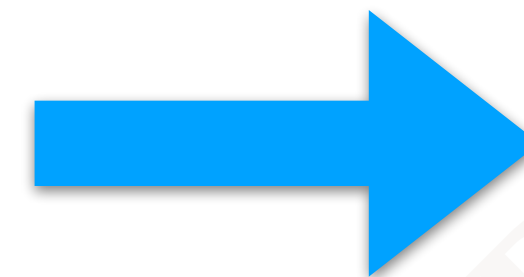
This may help explain why **LP-PRP** has shown superior *in vivo* results when administered to patients exhibiting signs of mild to moderate OA, due to its **PREDOMINANT ANTI-INFLAMMATORY** and **ANABOLIC** effects.

# BIOCHEMICAL EFFECTS OF PRP CONCENTRATES IN DIFFERENT TISSUES

**LP-PRP**



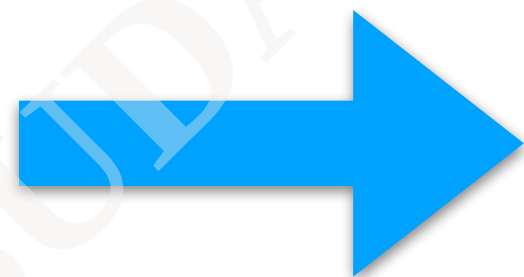
**ANABOLIC and ANTI-INFLAMMATORY**



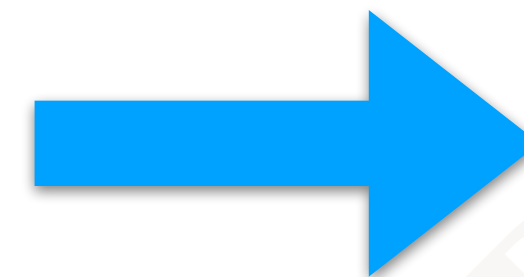
**MILD OSTEOARTHRITIS**

# BIOCHEMICAL EFFECTS OF PRP CONCENTRATES IN DIFFERENT TISSUES

**LR-PRP**



**CATABOLIC and PRO-INFLAMMATORY**



**MILD OSTEOARTHRITIS**



EXPERT OPINION ON BIOLOGICAL THERAPY  
<https://doi.org/10.1080/14712598.2020.1798925>



REVIEW



## Platelet-rich plasma for the treatment of knee osteoarthritis: an expert opinion and proposal for a novel classification and coding system

Elizaveta Kon<sup>a,b</sup>, Berardo Di Matteo<sup>a,b,c</sup>, Diego Delgado <sup>d</sup>, Brian J Cole<sup>e</sup>, Andrea Dorotei<sup>a,b</sup>, Jason L Dragoo<sup>f</sup>, Giuseppe Filardo<sup>g</sup>, Lisa A Fortier<sup>h</sup>, Alberto Giuffrida <sup>a,b</sup>, Chris H Jo<sup>i</sup>, Jeremy Magalon<sup>j,k</sup>, Gerard A Malanga<sup>l</sup>, Allan Mishra<sup>m</sup>, Norimasa Nakamura<sup>n</sup>, Scott A Rodeo<sup>o</sup>, Steven Sampson<sup>p</sup> and Mikel Sánchez <sup>d,q</sup>

**Table 2.** Explanation of the digits of the novel PRP coding system.

|                                     |                                       |                                                                                                                                                                                                                                                                                                                                  |
|-------------------------------------|---------------------------------------|----------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|
| <b>Number 1<br/>(N<sub>1</sub>)</b> | Basal platelet concentration in blood | 0 = 0–100,000 platelets/μL<br>1 = 100,000–200,000 platelets/μL<br>2 = 200,000–300,000 platelets/μL<br>3 = 300,000–400,000 platelets/μL<br>4 = 400,000–500,000 platelets/μL<br>5 = 500,000–600,000 platelets/μL<br>6 = 600,000–700,000 platelets/μL<br>7 = 700,000–800,000 platelets/μL<br>8 = 800 ...<br>9 = 900 ...<br>10 = ... |
| <b>Number 2<br/>(N<sub>2</sub>)</b> | Platelet concentration in PRP         | 0 = 0–100,000 platelets/μL<br>1 = 100,000–200,000 platelets/μL<br>2 = 200,000–300,000 platelets/μL<br>3 = 300,000–400,000 platelets/μL<br>4 = 400,000–500,000 platelets/μL<br>5 = 500,000–600,000 platelets/μL<br>6 = 600,000–700,000 platelets/μL<br>7 = 700,000–800,000 platelets/μL<br>8 = 800 ...<br>9 = 900 ...<br>10 = ... |
| <b>Number 3<br/>(N<sub>3</sub>)</b> | Red Blood Cells in PRP                | 0 = No presence/traces (<1x10 <sup>6</sup> /μL)<br>1 = Presence (>1x10 <sup>6</sup> /μL)                                                                                                                                                                                                                                         |
| <b>Number 4<br/>(N<sub>4</sub>)</b> | White Blood Cells in PRP              | 0 = Less than baseline<br>1 = 1.01 to 2 x baseline<br>2 = 2.01 to 3 x baseline<br>3 = 3.01 to 4 x baseline<br>4 = 4.01 to 5x baseline<br>5 = > 5x baseline                                                                                                                                                                       |
| <b>Number 5<br/>(N<sub>5</sub>)</b> | External Activation                   | 0 = No (endogenous)<br>1 = Yes                                                                                                                                                                                                                                                                                                   |
| <b>Number 6<br/>(N<sub>6</sub>)</b> | Calcium Addition                      | 0 = No<br>1 = Yes                                                                                                                                                                                                                                                                                                                |

### 3.6.2. Examples

(1) PRP obtained from blood with a mean concentration of 150,000 platelets/μL and reaching a mean concentration of 430,000 platelets/μL; presence of erythrocytes and no leukocytes; activation with thrombin: PRP code is 14–10–10.

(2) PRP obtained from blood with a mean concentration of 230,000 platelets/μL and reaching a mean concentration of 310,000 platelets/μL; no erythrocytes; double leukocyte concentration compared to blood levels and without exogenous activation: PRP code is 23–02–00.

(3) PRP obtained from blood with a mean concentration of 212,900 platelets/μL and reaching a mean concentration of 420,000 platelets/μL; with no leukocytes and no erythrocytes; and activation with calcium: PRP code is 24–00–11.

# CONTROVERSIES AND FUTURE DIRECTIONS OF RESEARCH

- 1) Is there a **DOSE-EFFECT RELATION** between the **PLATELET COUNT** and the **CLINICAL EFFICACY** of the preparation?
- 2) What is the role of platelet **ACTIVATION METHODS** on the **CLINICAL EFFICACY** of PRP concentrates?

## CONTROVERSIES AND FUTURE DIRECTIONS OF RESEARCH

- 3) Is there an **OPTIMAL NUMBER** of injections and **TIMEFRAME FOR APPLICATION** of multiple injection treatment cycles?
- 4) What is the **IMPACT** on clinical efficacy of opting for **FRESH VERSUS FREEZE-THAWED PRODUCTS**?

## LIST OF REFERENCES

- 1) Ferreira-Dos-Santos G. **From Spin Wizardry to Regenerative Alchemy: A Philosophical Inquiry Into Healing and Standardization.** *Reg Anesth Pain Med.* Published Online First: 29 April 2025.
- 2) Ferreira-Dos-Santos G, Hurdle MFB, Clendenen SR, Eldrige JS, Qu W. **Autologous Platelet-Rich Plasma Applications in Chronic Pain Medicine: Establishing a Framework for Future Research - A Narrative Review.** *Pain Physician.* 2022;25(1):15-27.
- 3) Navani A, Li G, Chrystal J. **Platelet rich Plasma in Musculoskeletal Pathology: A Necessary Rescue or a Lost Cause?** *Pain Physician.* 2017;20:E345-E356.

## LIST OF REFERENCES

- 4) Wasterlain AS, Braun HJ, Dragoo JL. **Contents and Formulation of Platelet-rich Plasma.** *Operative Tech Orthop.* 2012;22:33-42.
- 5) Kobayashi Y, Saita Y, Nishio H, *et al.* **Leukocyte Concentration and Composition in Platelet-rich Plasma (PRP) Influences the Growth Factor and Protease Concentrations.** *J Orthop Sci.* 2016;21:683-689.
- 6) Zhou Y, Zhang J, Wu H, Hogan MV, Wang JHC. **The Differential Effects of Leukocyte Containing and Pure Platelet-rich Plasma (PRP) on Tendon Stem/Progenitor Cells – Implications of PRP Application for the Clinical Treatment of Tendon Injuries.** *Stem Cell Res Ther.* 2015;6:173-186.

## LIST OF REFERENCES

- 7) Amaral RJ, da Silva NP, Haddad NF, *et al.* **Platelet-rich Plasma Obtained with Different Anticoagulants and their Effect on Platelet Numbers and Mesenchymal Stromal Cells Behavior in Vitro.** *Stem Cells Int.* 2016;2016:7414036.
- 8) Castillo TN, Pouliot MA, Kim HJ, Dragoo JL. **Comparison of Growth Factor and Platelet Concentration from Commercial Platelet-rich Plasma Separation Systems.** *Am J Sports Med.* 2011;39(2):266-271.
- 9) Marx RE. **Platelet-rich Plasma (PRP): What is PRP and what is not PRP?** *Implants Dental.* 2001;10:225-228.

## LIST OF REFERENCES

- 10) Weibrich G, Hansen T, Kleis W, Buch R, Hitzler WE. **Effect of Platelet Concentration in Platelet-rich Plasma on Peri-implant Bone Regeneration.** *Bone*. 2004;34:665-671.
- 11) Assirelli E, Filardo G, Mariani E, *et al.* **Effect of Two Different Preparations of Platelet-rich Plasma on Synoviocytes.** *Knee Surg Sports Traumatol Arthrosc*. 2015; 23:2690-2703.
- 12) Knezevic NN, Candido KD, Desai R, Kaye AD. **Is Platelet-rich Plasma a Future Therapy in Pain Management?** *Med Clin North Am*. 2016;100:199-217.



**QUESTIONS/COMMENTS?**  
**THANK YOU!**

**Presenter:** Guilherme (Gui) Dos Santos, MD | **Department of Anesthesiology, Reanimation, and Pain Medicine**

Hospital Clínic de Barcelona, University of Barcelona